## Concise Total Syntheses of Aspalathin and Nothofagin

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Received February 5, 2010

## ABSTRACT

BnO<sup>OT</sup> OBn X = H or OBn X = OBn OH OH OH X = H: Nothofagin X = OH: Aspalathin

Syntheses of the *C*-glycosyl flavone natural products aspalathin and nothofagin have been accomplished in eight steps from tribenzyl glucal, tribenzylphloroglucinol, and either 4-(benzyloxy)phenylacetylene or 3,4-bis(benzyloxy)phenylacetylene. The key step of the syntheses involves a highly stereoselective Lewis acid promoted coupling of 1,2-di-*O*-acyl-3,4,6-tribenzylglucose with tribenzylphloroglucinol, which gives rise to the corresponding  $\beta$ -*C*-aryl glycoside in 30–65% yields.

Naturally occurring C-aryl glycosides exhibit a range of interesting biological properties. C-Glycosyl flavonoids, in particular, have been shown to possess antiviral, cytotoxic, and DNA binding activities.<sup>1</sup> The natural product aspalathin, isolated from the leaves of Aspalathus linearis and used in the manufacture of rooibos tea,<sup>2</sup> displays potent antioxidant and radical scavenging activity<sup>3</sup> and has recently been found to inhibit proliferation and infiltration of liver cancer cells.<sup>4</sup> The structurally related flavonoid nothofagin, typically coisolated with aspalathin,<sup>2</sup> also displays antioxidant properties, but to a lesser extent than aspalathin. Both of these compounds can be considered as direct precursors of a variety of other biologically interesting C-aryl flavonoid natural products, such as orientin/isoorientin,<sup>5</sup> vitexin/isovitexin,<sup>5</sup> and aspalalinin<sup>6</sup> (Figure 1). In view of their impressive biological profile, we decided to undertake total syntheses of both aspalathin and nothofagin.

Structurally, both natural products consist of a glucopyranosyl unit carbon-linked to a dihydrochalcone moiety, with  $\beta$ -stereochemistry observed at the anomeric carbon atom.<sup>2</sup> We envisioned that the crucial linkage between the carbohydrate and aromatic subunits could be fashioned by a Lewisacid promoted Friedel–Crafts-type glycosylation reaction; this process is well-precedented in the literature when highly electron-rich aromatics are employed as nucleophiles,<sup>7,8</sup> and thus this method seemed a logical starting point for our investigations.

Addition of TESOTF (1 equiv) to a solution of tribenzylphloroglucinol (**2a**,<sup>9a</sup> 2 equiv) and 2,3,4,6-tetra-*O*-benzylglucose-1-*O*-acetate<sup>9b</sup> (**1**) in CH<sub>2</sub>Cl<sub>2</sub> at 0 °C resulted in a clean conversion to the corresponding *C*-aryl glycoside as a ~3:1 mixture of  $\beta/\alpha$  diastereomers in 89% yield (**3a** and **3b**, Scheme 1). Indeed, Schmidt also previously obtained 3:1 diastereoselectivity in the ZnCl<sub>2</sub>-promoted coupling of 2,3,4,6-tetrabenzylglucose-1-*O*-trichloroacetimidate with

<sup>(1)</sup> Furuta, T.; Kimura, T.; Kondo, S.; Mihara, H.; Wakimoto, T.; Nakaya, H.; Tsuji, K.; Tanaka, K. *Tetrahedron Lett.* **2004**, *60*, 9375.

<sup>(2)</sup> Koeppen, B. H.; Roux, D. G. Biochem. J. 1966, 99, 604.

<sup>(3)</sup> von Gadow, A.; Joubert, E.; Hansmann, C. F. J. Agric. Food. Chem. **1997**, 45, 632.

<sup>(4)</sup> Snijman, P. W.; Swanevelder, S.; Joubert, E.; Green, I. R.; Gelderblom, W. C. A. *Mutat. Res., Genet. Toxicol. Environ. Mutagen.* **2007**, *631*, 111.

<sup>(5)</sup> Krafczyk, N.; Glomb, M. A. J. Agric. Food Chem. 2008, 56, 3368.
(6) Shimamura, N.; Miyase, T.; Umehara, K.; Warashina, T.; Fujii, S. Biol. Pharm. Bull. 2006, 29, 1271.

<sup>(7)</sup> Stewart, A. O.; Williams, R. M. J. Am. Chem. Soc. 1985, 107, 4289.

<sup>(8)</sup> Schmidt, R. R.; Hoffmann, M. Tetrahedron Lett. 1982, 23, 409.

<sup>(9) (</sup>a) Kawamoto, H.; Nakatsubo, F.; Murakami, K. Synth. Commun. **1996**, 26, 531. (b) 2,3,4,6-Tetra-O-benzylglucose-1-O-acetate was prepared from dextrose in 60% overall yield by allylation (allyl alcohol, H<sub>2</sub>SO<sub>4</sub>), benzylation (NaH, BnBr, DMF), deallylation (*t*-BuOK, DMF, 70 °C; 1 N H<sub>2</sub>SO<sub>4</sub>), and acylation (Ac<sub>2</sub>O, Pyr) . (c) Prepared in 70% yield by treatment of phloroacetophenone with benzyl chloride (3.3 equiv) and K<sub>2</sub>CO<sub>3</sub> (5 equiv) in DMF at 80 °C for 1 h.



**Figure 1.** Aspalathin and nothofagin as precursors of other naturally occurring C-glycosyl flavones.

**2a**.<sup>10</sup> However, attempted coupling of 2,4,6-tris(benzyloxy)acetophenone (**2b**,<sup>9c</sup> 2 equiv) and **1** in the presence of TESOTF (1–5 equiv) gave only trace quantities (<5%) of the desired *C*-aryl glycoside product, and unreacted starting materials were recovered in high yield.

To circumvent the low diastereoselectivity obtained in the coupling reaction, we explored an alternative approach based on the *C*-glycosidation studies of Kishi<sup>11</sup> involving anchimeric participation by a C2 protecting group.

Seeking to avoid the formation of ketal byproduct observed in reactions involving C2 acetate-protected carbohydrates (ie., 6,  $R = CH_3$ , Scheme 1), we first prepared 1,2-di-O-pivaloyl-3,4,6-tribenzylglucose 4a from tribenzyl glucal by Oxonemediated dihydroxylation,<sup>12</sup> followed by treatment with excess trimethylacetyl chloride in pyridine (60 °C, 24 h). Reaction of 4a with tribenzylphloroglucinol (2a, 4 equiv) at 0 °C in 10:1 CH<sub>2</sub>Cl<sub>2</sub>/THF in the presence of 10 equiv of TMSOTf afforded glycoside 5 as a single diastereomer in 55% yield. The  $\beta$ -stereochemistry of the carbohydrate was confirmed in the 9.2 Hz coupling constant observed for the carbohydrate C2 proton. As anticipated, reaction of 4a with 2b under these conditions gave no reaction. Compound 5 could be transformed into 3a by DIBAL reduction followed by benzylation of the C2 alcohol (KH, BnBr, THF) in 80% overall yield.

Although a stereoselective glycosylation reaction had been found, the moderate yields of product obtained, along with the excess of nucleophile and promoter required, indicated the significantly lower reactivity of carbohydrate **4a** as compared to **1**. We therefore performed an optimization study by altering the reaction promoter, as well as the identity of the acyl protecting groups (R) on glycosyl acceptor **4** (Table

Scheme 1. C-Glycosylation of Carbohydrates 1 and 4a



1). Due to the sparing solubility of **2a** in acetonitrile and ether at 0 °C, solvent systems chosen for the reaction were admixtures of CH<sub>2</sub>Cl<sub>2</sub> with acetonitrile or THF; reactions performed in  $CH_2Cl_2$  alone were slow (t > 1 h), and the optimal solvent combination found was 2:1 CH<sub>2</sub>Cl<sub>2</sub>/THF. Of the Lewis acids surveyed, only TMSOTf, TESOTf, and SnCl<sub>4</sub> (entries 1, 2, 4, 5, 8, and 9) gave rise to the desired product in appreciable yields; the use of BF<sub>3</sub>•OEt<sub>2</sub> (entry 3), InCl<sub>3</sub>, (entry 6), or Et<sub>2</sub>AlCl (entry 7) gave negligible amounts of 5 even after prolonged reaction times (>24 h) at ambient temperature. For glycosyl acceptors 4a and 4c, excess promoter ( $\geq 10$  equiv) was required for optimal conversions of the starting material. Furthermore, in all cases, highest yields of  $5^{13}$  were obtained when 3.5–4.0 equiv of tribenzylphloroglucinol was employed; unreacted 2a could be efficiently recovered (>2 equiv) after column chromatography of the crude reaction mixture.

Le Drian<sup>14</sup> has suggested that the use of the less bulky isobutyryl ester as a carbohydrate protecting group for glycosidation reactions involving anchimeric participation affords higher product yields than similar reactions employing pivaloate-protected sugars. Indeed, coupling of bisisobutyrate **4b**<sup>15a</sup> with **2a** (4 equiv) in the presence of 5 equiv of TMSOTf led to **5b** in 65% yield (86% based on recovered starting material); ketal byproduct **6** (R = *i*Pr) was not formed to any significant extent (<5%) in the reaction. However, reaction of cyclohexyl ester **4c**<sup>15b</sup> with **2a** (4 equiv) proceeded sluggishly, providing ~30% **5c** after the addition

<sup>(10)</sup> Schmidt, R. R.; Effenberger, G. Carbohydr. Res. 1987, 171, 59.
(11) Minehan, T. G.; Kishi, Y. Tetrahedron Lett. 1997, 38, 6815.

<sup>(12)</sup> Rani, S.; Vankar, Y. D. *Tetrahedron Lett.* **2003**, 44, 907. While a  $\sim$ 1:1 mixture of anomers at C1 of the carbohydrate is observed in the

dihydroxylation reaction, the diastereoselectivity at C2 of the carbohydrate is observed in the disydroxylation reaction, the diastereoselectivity at C2 of the carbohydrate is very high, and we estimate a >95:5 diastereomer ratio at C2 by <sup>1</sup>H NMR analysis of the individual  $\alpha$  and  $\beta$  C1 anomers of dipivaloate **4a**.

<sup>(13)</sup> Attempts to achieve greater conversions in the *C*-glycosylation reactions by adding excess Lewis acid (>10 equiv), by elevating the reaction temperature ( $\rightarrow$ 25 °C), or by increasing the reaction time (>1h) only led to lower overall yields of **5**, presumably due to acid-induced cleavage of the aromatic benzyl ether protecting groups.

<sup>(14)</sup> Desmares, G.; Lefebvre, D.; Renevret, G.; Le Drian, C. Helv. Chim. Acta 2001, 84, 880.

<sup>(15) (</sup>a) Prepared in the same manner as 4a, except with the substitution of isobutryl chloride for pivaloyl chloride in the esterificaton step. (b) Prepared in the same manner as 4a, except with the substitution of cyclohexyl carbonyl chloride for pivaloyl chloride in the esterificaton step.





<sup>*a*</sup> Optimal yields were obtained using 3.5-4.0 equiv of **2a**. The optimal reaction concentration was 0.33 M relative to **4**. For entries 1-8, <5% ketal byproduct **6** was formed, as assayed by <sup>1</sup>H NMR of the crude reaction mixture. <sup>*b*</sup> Yield in parentheses represents yield based on recovered starting material.

of ~12 equiv of TMSOTf (entry 8); **4c** was recovered in 45% yield from the reaction after workup. Reaction of diacetate **4d**<sup>12</sup> with **2a** in the presence of 5 equiv of TMSOTf gave a 43% yield of **5d**, along with ~35% recovered starting material and 12% of an unidentified byproduct presumably derived from acetal **6** ( $\mathbf{R} = \mathbf{CH}_3$ ).

It was originally envisioned that benzyl ether 3a (efficiently prepared from 5a, Scheme 1) could be advanced toward the natural products by introduction of a carbonyl substituent on the phloroglucinol aromatic ring. Friedel-Crafts acylation approaches employing acetyl chloride/AlCl<sub>3</sub> or acetic anhydride in combination with trifluoroacetic acid,<sup>16</sup> phosphoric acid,<sup>17</sup> zinc chloride,<sup>18</sup> indium chloride,<sup>19</sup> or indium triflate<sup>20</sup> all led to substrate decomposition due to premature loss of the aromatic benzyl ether protecting groups under the acidic reaction conditions. We ultimately found, however, that Vilsmeier-Haack reaction<sup>21</sup> of **3a** (DMF, POCl<sub>3</sub>) smoothly led to aldehyde 7a in 90% yield (Scheme 2). Attempts to repeat this reaction with N.N-dimethylacetamide in place of DMF (to furnish the corresponding aryl ketone) at room temperature or at elevated temperatures gave no reaction. Analysis of the <sup>1</sup>H NMR spectrum of compound 7a revealed the presence of two diastereomers in a ratio of 2:1. From this observation, we concluded that anomerization was occurring under the Vilsmeier reaction conditions to give a thermodynamic mixture of  $\beta$ - and  $\alpha$ -C-aryl glycosides. To avoid compromise of the anomeric stereochemistry in this manner, we chose to investigate the direct formylation of glycoside **5a** containing the C2 pivaloyloxy protecting group.<sup>22</sup> Fortuitously, aldehyde **7b** was obtained in 85% yield with a 10:1 diastereomer ratio in favor of the desired  $\beta$ -*C*-aryl glycoside.





For completion of the natural product syntheses we envisioned addition of an appropriate alkynyllithium species to aldehyde 7b, followed by benzylic oxidation and protecting group removal. The requisite alkynes 10a<sup>23</sup> and 10b for this procedure were fashioned from 4-hydroxybenzaldehyde 8a and 3,4-dihydroxybenzaldehyde 8b by benzylation<sup>24</sup> and alkynylation with the Bestmann–Ohira reagent (Scheme 3).<sup>25</sup> Treatment of alkyne 10a with 0.9 equiv of *n*-BuLi, followed by addition of aldehyde **7b** gave rise to a  $\sim$ 1:1 mixture of diastereomeric propargylic alcohols (11a and 11b, 81%) which was immediately treated with CH<sub>3</sub>MgBr in ether to effect removal of the C2 pivaloyl protecting group. Oxidation of diols 12a,b with excess MnO<sub>2</sub> in 1:1 CH<sub>2</sub>Cl<sub>2</sub>/hexanes gave rise to ynone 13 in 93% overall yield from 11a,b. Finally, hydrogenolysis of the benzyl ether protecting groups ( $H_2$ , 10% Pd on C, rt, 18 h) gave an 89% yield of synthetic nothofagin, the spectrocopic and physical data of which (<sup>1</sup>H and <sup>13</sup>C NMR, UV, and melting point) matched those reported for the natural compound.<sup>5,26</sup>

The synthesis of aspalathin was accomplished in an analogous manner (Scheme 4) by coupling lithiated **10b** with

<sup>(16)</sup> Nay, B.; Arnaudinaud, V.; Vercauteren, J. Eur. J. Org. Chem. 2001, 12, 2379.

<sup>(17)</sup> Gjoes, N.; Gronowitz, S. Acta Chem. Scand. 1972, 26, 1851.

<sup>(18)</sup> Kawamoto, H.; Nakatsubo, F.; Murakami, K. J. Wood Chem. Technol. 1989, 9, 35.

<sup>(19)</sup> Hayashi, R.; Cook, G. R. Org. Lett. 2007, 9, 1311.

<sup>(20)</sup> Koshima, H.; Kubota, M. Synth. Commun. 2003, 33, 3983.

<sup>(21)</sup> Jones, A. W.; Wahyuningsih, T. D.; Pchalek, K.; Kumar, N.; Black, D. *Tetrahedron* **2005**, *61*, 10490.

<sup>(22)</sup> Glycoside **5b** could also be used in the Vilsmeier formylation reaction, providing an 83% yield of the corresponding isobutyryl-protected aryl aldehyde. This substrate was not chosen to advance through the remainder of the synthesis for two reasons: first, the cost of isobutyryl chloride required for the preparation of **4b** (and thus **5b**) is about twice that of pivaloyl chloride, and second, slightly lower overall yields ( $\sim$ 55%) were obtained when the isobutyryl-protected aryl aldehyde was subjected to the coupling (with lithiated **10a** and **10b**) and isobutyrate deprotection reactions.

<sup>(23)</sup> Maehr, H.; Uskokovic, M. R.; Schaffner, C. P. Synth. Commun. 2009, 39, 299.

<sup>(24) 4-</sup>Benzyloxybenzaldehyde: Narasimhulu, M.; Srikanth Reddy, T.; Chinni Mahesh, K.; Sai Krishna, A.; Venkateswara Rao, J.; Venkateswarlu, Y. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 3125. 3,4-Bis(benzyloxy)benzaldehyde: Milhazes, N.; Cunha-Oliveira, T.; Martins, P.; Garrido, J.; Oliviera, C.; Rego, A.; Borges, F. *Chem. Res. Toxicol.* **2006**, *10*, 1294.

 <sup>(25) (</sup>a) Muller, S.; Liepold, B.; Roth, R. G.; Bestmann, H. J. Synlett
 1996, 521. (b) Ohira, S. Synth. Commun. 1989, 19, 561.

<sup>(26)</sup> Melting point and UV data for nothofagin: Hillis, W. E.; Inoue, T. *Phytochemistry* **1967**, *6*, 59. To the best of our knowledge, there exist in the literature no reports of the measurement of the specific rotation ( $[\alpha]_D$ ) of nothofagin.

Scheme 3. Completion of the Synthesis of Nothofagin



**7b**, followed by pivaloyl deprotection, benzylic oxidation, and benzyl ether hydrogenolysis; synthetic aspalathin was obtained in 51% overall yield from **14a**,**b**. The spectroscopic and physical data for the synthetic material (<sup>1</sup>H and <sup>13</sup>C NMR, UV, IR,  $[\alpha]_D$ , and melting point) was identical in all respects to those reported for the natural product.<sup>2,5</sup>

In summary, eight-step syntheses of the natural glycosyl flavonoids nothofagin and aspalathin have been accomplished in 28% and 20% overall yield, respectively, from tribenzyl glucal, tribenzylphloroglucinol, and either 4-(benzyloxy)phenylacetylene or 3,4-bis(benzyloxy)phenylacetylene. We are currently investigating the oxidative transformation of these Scheme 4. Completion of the Synthesis of Aspalathin



compounds into other products of potential medicinal and therapeutic value.

Acknowledgment. We thank the National Institutes of Health (SC2 GM081064-01), the ACS Petroleum Research Fund (No. PRF45277-B1), and the Henry Dreyfus Teacher—Scholar Award for their generous support of our research program. Mass spectra were provided by the UC Riverside Mass Spectrometry facility.

**Supporting Information Available:** Detailed experimental procedures, spectroscopic data, and <sup>1</sup>H and <sup>13</sup>C NMR spectra for compounds in Table 1 and Schemes 1–4. This material is available free of charge via the Internet at http://pubs.acs.org.

OL100315G